# RESIN ACIDS OF PINUS RESINOSA NEEDLES

## DUANE F. ZINKEL and WILBUR B. CLARKE\*

Forest Products Laboratory,† P.O. Box 5130, Madison, WI 53705, U.S.A.; \*Southern University, Department of Chemistry, Southern Branch, P.O. Box 9797, Baton Rouge, LA 70813, U.S.A.

(Received 10 September 1984)

Key Word Index-Pinus resinosa; Pinus nigra; Pinaceae; needles; labdane diterpenes; hybrids.

Abstract—The principal resin acids in the needles of *Pinus resinosa* are the labdane diterpenes, the new 8,13-epoxy-14-labden-19-oic acid (epimanoyl oxide acid), 8,13 $\beta$ -epoxy-14-labden-19-oic acid (manoyl oxide acid), 8(17),E-12,14-labdatrien-19-oic acid (communic acid) and 15-oxo-8(17)-labden-19-oic acid (imbricataloic acid). A survey of needles from representative populations of *P. resinosa* showed a limited variability in resin acid composition consistent with the uniformity of other traits. The composition of needle resin acids for putative *P. nigra* × resinosa hybrids strongly suggests the improbability of *P. resinosa* as the pollen parent.

### INTRODUCTION

American red pine, *Pinus resinosa* Ait., is an important North American pulpwood and lumber conifer in the Great Lakes States and Canada. Historically, recovery of naval stores from the oleoresin has been nearly non-existent, the only practical use having been as a water-proof sealer for birch bark canoes [1]. Naval stores, however, are recovered from red pine wood as byproducts of kraft pulping.

As part of an evaluation of pine foliage as a potential source for naval stores and fine chemicals, we have investigated the diterpene resin acids of red pine needles. This paper reports on the unique composition of resin acids in red pine needles, on the isolation of two new labdane resin acids, and on specific implications of needle resin acid composition in genetic studies.

#### RESULTS AND DISCUSSION

Preliminary GLC analysis of *P. resinosa* needle resin acids showed the major components to be different both in kind and quantity from the common tricyclic pimaric, isopimaric and abietadienoic acids found in the xylem of this [2-4] and many other pines. These common resin acids are, however, only about 10% of the total in the needles; four other resin acids, all labdane diterpenes, constitute the major portion.

Identification of resin acids

The resin acid fraction of P. resinosa needle ether extract was crystallized from ether-petrol. Recrystal-

lization gave a pure acid which was converted to the methyl ester. Argentative CC of the methylated acids from the acid crystallization-liquors provided a second resin acid methyl ester similar to the above methyl ester in its spectral characteristics. Spectral data for the two esters (M, 334) suggested that they were C-19 carbomethoxy (chemical shift for C-10 hydrogens at about  $\delta$ 0.6) manoyl oxide/epimanoyl oxide derivatives. Comparison of the <sup>1</sup>H NMR patterns of the vinyl ABX hydrogens with those for manoyl oxide and epimanoyl oxide strongly suggested the equatorial vinyl configuration (i.e. the methyl ester, 1b, of manoyl oxide acid,  $8,13\beta$ -epoxy-14-labden-19-oic acid, 1a) for the acid obtained by crystallization, and the axial vinyl configuration for the methyl ester, 2b (of epimanoyl oxide acid, 8,13-epoxy-14-labden-19-oic acid, 2a) obtained by argentative CC (Table 1). A stronger silver complexing of 1b (i.e. greater chromatographic retention) than 2b further supports these assignments. We have also found 1a in the needles of P. nigra [5, 6] and P. sylvestris [5]. Since completion of this work, Bardyshev et al. have reported the isolation and identification of 1a from P. sylvestris needles [7]. The enantiomeric, C-18 epimanoyl oxide acid has been isolated from a Beyeria species [8] and three double-bond isomers of epimanool-19-oic acid have been isolated from Agathis robusta [9].

The two other major labdane resin acids were isolated as the methyl esters by argentative chromatography and identified as methyl communate (methyl 8(17),E-12,14-labdatrien-19-oate, 3) and methyl imbricataloate (methyl 15-oxo-8(17)-labden-19-oate, 4). Communic acid (communic acid ≡ elliotinoic acid [10]) has been reported to be present in small amounts in the xylem oleoresins of P. elliottii, P. edulis and P. strobus [11] and in the cortex oleoresin of P. strobus [12]. Imbricataloic acid has been isolated from the needles of P. elliottii [13].

Composition of P. resinosa needle resin acids

A survey of the needle resin acid composition of *P. resinosa* was conducted on samples of nine populations representing the major habitats of the species. The

<sup>†</sup> Maintained by the Forest Service of the U.S. Department of Agriculture, in cooperation with the University of Wisconsin.

The use of trade, firm, or corporation names in this publication is for the information and convenience of the reader. Such use does not constitute an official endorsement or approval by the U.S. Department of Agriculture of any product or service to the exclusion of others which may be suitable.

Table 1. <sup>13</sup>C chemical shifts of manoyl oxide acid methyl ester 1b, the dihydro derivative 1c, and epimanoyl oxide acid methyl ester 2b

Carbon atom	1 <b>b</b> δ	lc δ	<b>2b</b> δ	Carbon atom	1 <b>b</b> δ	1c δ	<b>2b</b> δ
1	39.4	39.5	39.8	11	15.5	15.6	16.2
2	19.1	19.2	19.3	12	35.5	35.9	35.1
3	38.2	38.2	38.3	13	73.2	72.8	73.2
4	43.9	43.9	44.0	14	148.0	37.9	147.8
5	55.0*	56.9*	57.0*	15	110.1	7.9	109.6
6	21.5	21.4	21.5	16	28.8	27.1	32.7
7	43.3	43.0	43.2	17	25.0	24.3	23.5
8	74.7	74.1	75.7	18	28.7	28.5	28.6
9	57.0*	57.8*	58.0*	19	177.5	177.6	177.6
10	37.5	37.2	37.4	20	13.0	13.2	13.4
				OMe	51.0	51.0	51.0

<sup>\*</sup>Chemical shift assignments between C-5 and C-9 and between C-16 and C-17 are interchangeable.

1a R = H

1b R = Me

1c R = Me; vinyl reduced to Et

2a R = H

2b R = Me

$$\mathbb{R}^2$$
  $\mathbb{R}^1$   $\mathbb{R}^2$ 

R<sup>1</sup> R<sup>2</sup> **3** Me CO<sub>2</sub>Me **5** CO<sub>2</sub>Me Me

$$R^{2}$$
 $R^{1}$ 
 $R^{2}$ 
 $R^{2$ 

Me

6 CO₂Me

methylated resin acids were analysed by packed column GC. The similar retention characteristics for several of the components required analysis on two columns, one containing DEGS and the other SE-30/EGiP [14]; decreasing column temperature from the usual 200 to 186° improved resolution (Table 2).

The needle resin acid composition (Table 3) is relatively consistent among the nine *P. resinosa* populations studied, in agreement with the uniformity in characteristics of the species [16, 17]. The variations that are present correspond fairly well with patterns of latitudinal

variation reported for other traits [Lester, D. T., private communication]. Thus, the more southerly provenances 703, 761, and 774 are lower in manoyl oxide/epimanoyl oxide acids (and total oxygenated labdane acids 1a, 2a, and 4) and higher in communic acid than the other more northerly provenances examined.

The above provenance data for resin acid composition represented mature needles of trees from a 10-year-old plantation. Because early characterization is invaluable in genetic and chemotaxonomic studies, we also examined the consistency of resin acid composition in young needle

Table 2. GC retention data\* for the methyl esters of the major resin acids in P. resinosa needles

	DE	GS	SE-30/EGiP		
Resin acid (as methyl ester)	186°	200°	186°	200°	
Manoyl oxide acid, 1a	1.27	1.19	0.98	0.97	
Epimanoyl oxide acid, 2a	1.26	1.24	1.06	1.04	
Communic acid, 3	1.36	1.28	1.08	1.05	
4-Epicommunic acid	1.69	1.53	1.22	1.18	
Imbricataloic acid, 4	3.8	3.63	1.56	1.58	

<sup>\*</sup>Retention relative to methyl pimarate = 1.00.

tissue and in young plants. Primary needles of the previous year from 2-year-old seedlings (September sampling) contained only the usual tricyclic resin acids and none of the labdane resin acids, but the secondary needles

of current year's growth were high in communic acid (3) and still comparatively low in the manoyl oxide/epimanoyl oxide acids (1a, 2a). For 3-year-old seedlings, the manoyl oxide/epimanoyl oxide acid content had increased considerably but the content in the current year's needles was still low (total ca 20%). By a tree age of 5 years, however, resin acid content in needles of each of the previous two growing seasons was consistent and in line with values observed for older trees. New-growth needles, however, had a lower content of manoyl oxide/epimanoyl oxide acids similar to observations for manoyl oxide acid in certain P. nigra trees [5]. Thus, comparative studies of resin acid composition of P. resinosa needles should use only mature needles from plants at least 5 years old.

The resin acids in the cortex oleoresin of several trees were analyzed for comparison with those in the needles. The cortical resin acids of *P. resinosa* are characterized by a high content of levopimaric/palustric, neoabietic and communic acids; manoyl oxide/epimanoyl oxide acids were not observed.

Table 3. Composition of needle resin acids of Pinus resinosa

	Percent of total resin acids†							
Source*	Sand	MOA	EMOA	Com	L/P	Neo	Imb	
703 Gladwin Co., Michigan	2.2	32.0	7.2	39.0	5.0	4.0	10.3	
761 Essex Co., New York	1.8	35.6	11.4	32.5	6.5	3.6	8.3	
774 Langlade Co., Wisconsin	1.6	29.6	7.6	45.8	3.2	1.3	10.4	
724 Queens Co., New Brunswick	1.1	49.1	14.2	23.4	3.1	3.2	5.6	
735 Marquette Co., Michigan	2.4	46.5	11.8	23.7	3.6	2.3	9.2	
752 Berthier Co., Quebec	1.9	49.8	14.5	17.7	4.8	2.5	8.6	
755 Itaska Co., Minnesota	1.0	50.8	12.0	21.3	2.2	2.0	10.5	
777 Douglas Co., Wisconsin	1.6	48.1	12.2	23.2	4.0	4.4	6.3	
781 Nipissing Co., Ontario	1.1	50.0	7.5	25.7	2.8	0.9	11.6	

<sup>\*</sup>Needles of the previous year were obtained in April 1973 and were a composite from two branches of each of three trees. The trees were from a 10-year-old experimental plot in Central Wisconsin. Provenance (source) designations (numbers) per [15].

Table 4. Composition of needle resin acids in P. nigra x resinosa hybrids

Percent of total resin acids†							
Sand	MOA	L/P	Ecom	Ab	DeAb	Neo	
3.2	_	23.3	31.0	3.3	7.3	21.4	
5.9		14.6	62.8	3.0	4.2	7.1	
5.0	39.3	14.9	30.7	1.3	2.6	4.4	
4.4	46.0	18.5	16.6	1.7	6.0	6.7	
	3.2 5.9 5.0	3.2 — 5.9 — 5.0 39.3	Sand         MOA         L/P           3.2         —         23.3           5.9         —         14.6           5.0         39.3         14.9	Sand         MOA         L/P         Ecom           3.2         —         23.3         31.0           5.9         —         14.6         62.8           5.0         39.3         14.9         30.7	Sand         MOA         L/P         Ecom         Ab           3.2         —         23.3         31.0         3.3           5.9         —         14.6         62.8         3.0           5.0         39.3         14.9         30.7         1.3	Sand         MOA         L/P         Ecom         Ab         DeAb           3.2         —         23.3         31.0         3.3         7.3           5.9         —         14.6         62.8         3.0         4.2           5.0         39.3         14.9         30.7         1.3         2.6	

<sup>\*</sup>Specific hybrid number per Dr W. B. Critchfield. The trees were 15 years old when sampled.

<sup>†</sup>By calculation of GC data from SE-30/EGiP and DEGS packed columns [14] at 186°. Sand = sandaracopimaric, MOA = manoyl oxide acid (1a), EMOA = epimanoyl oxide acid (2a), Com = communic (3), L/P = levopimaric/palustric (unresolved), Neo = neoabietic, Imb = imbricataloic (4). Values for Imb are the sum of imbricataloic and an artifact methyl ketone [13] peaks. Small amounts of abietic and dehydroabietic acids were present in all samples.

<sup>†</sup>Ecom = epicommunic acid, Ab = abietic acid and DeAb = dehydroabietic acid. Other abbreviations and GC conditions as per Table 1. Hybrid No. 11 contains about 5% each of pinifolic and 4-epiimbricataloic [6] acids; the three other hybrids contained lesser amounts. GC data for hybrid × hybrid and hybrid × nigra back crosses are available from the senior author.

Composition of needle resin acids in P. resinosa hybrids

Attempts at genetic improvement in red pine through interspecific hybridization have met with only limited success. Critchfield reported [18] a P. nigra × resinosa cross, albeit only four progeny were selected by foliage characteristics as true hybrids. (A recent publication of Gambles and Dengler [19] provides a detailed description of the morphology of P. resinosa needles.) The availability of plant material from these hybrids provided an opportunity to further examine diterpene resin acid composition as a tool for the characterization of hybrids.

The needle resin acid composition for the four P. nigra × resinosa putative hybrids is given in Table 4. Although the P. resinosa parent was no longer living, the limited variability in the species, as seen in Table 1, permits a general needle resin acid characterization of the parent as 30-50 % manoyl oxide acid, 7-14 % epimanoyl oxide acid, 18-46% communic acid and 6-12% imbricataloic acid. Although the composition of P. nigra needle resin acids varies widely [6], the resin acids of the hybrids' P. nigra parent consisted primarily of manoyl oxide acid (63%) with a total of 11% of epimbricataloic and pinifolic acids (P. nigra N1-N7, [6]); epimanoyl oxide acid and communic acid were not present. Because the needles of both parents are characterized by a high content of manoyl oxide acid 1a, it was surprising that, while two of the progeny have about 40% la in the needle resin acids, two progeny did not contain any. Equally puzzling was the presence of a resin acid, 4-epicommunic acid, 5, not found in either of the hybrids' parents. Epicommunic acid is the major needle resin acid of Spanish P. nigra [6] and P. densiflora [20]. The needle resin acid composition of the four hybrids argues against P. resinosa as the pollen parent in the crosses. Rather, a contaminated pollen resulting in some other P. nigra hybrid such as P. nigra × densiflora appears more probable. This is supported by a re-examination of the hybrids based on enzymespecifying loci [21].

## **EXPERIMENTAL**

GC was done with a Hewlett-Packard model 5750 gas chromatograph equipped with FID. <sup>1</sup>H NMR and <sup>13</sup>C NMR were recorded at 250 and 62.89 MHz, respectively, in CDCl<sub>3</sub> with TMS as int. standard. Mps were determined in evacuated glass capillaries and are corrected. Optical rotations were done using chloroform as solvent.

Isolation and identification of resin acids. Fresh needles of Pinus resinosa were shredded and extracted with Et<sub>2</sub>O. The volume of Et<sub>2</sub>O was reduced and 2 parts petrol added. After refrigeration overnight, the solution was filtered and then extracted with 1 N NaOH. The NaOH solution was washed 2 times with petrol, acidified with HCl and the resulting turbid solution extracted with petrol. The petrol solution was reduced in volume and refrigerated to give a precipitate. Recrystallization of the precipitate from methanol and then from pentane gave pure 1a (as shown by GC of the methyl ester): mp 147-147.5° (lit. [7] gives an unexplainably low value of 64-66°);  $[\alpha]_D^{25}$  -41.5° (c 0.9), (lit. [7], -39.2°, EtOH); (Found: C, 74.84; H, 10.10. C<sub>20</sub>H<sub>32</sub>O<sub>5</sub> requires C, 74.97; H, 10.07%). Methylation with diazomethane gave methyl ester 1b: mp 84.3-84.5° (lit. [7], 83-85°);  $[\alpha]_D^{25}$  - 47.6° (c1.2), (lit. [7],  $-43.2^{\circ}$ , EtOH): <sup>1</sup>H NMR:  $\delta$ 0.62 (Me-20), 1.17 (Me-18), 1.27 (Me-16), 1.30 (Me-17), 4.91 (1H, dd, J = 1.5 and 10.7 Hz), 5.14 (1H, dd, J = 1.5 and 17.5 Hz), and 5.87 (1H, dd, J= 10.7 and 17.5 Hz); chemical shifts in CCl<sub>4</sub> were 0.02-0.04 further downfield than reported previously [7]. IR and MS were consistent with those reported by Bardyshev *et al.* [7] with the exception of m/z at 287, 236, and 189 rather than 288, 237, and 190.  $^{13}$ C NMR is given in Table 1 (cf. [7]).

Compound 1b was hydrogenated at ambient conditions in benzene using triphenylphosphine rhodium chloride catalyst. Recrystallization from MeOH-H<sub>2</sub>O gave the hydrogenated derivative, methyl 8,13 $\beta$ -epoxy-19-labdanoate 1c: mp 51-52°; [ $\alpha$ ] $_{25}^{25}$  - 34.3° (c0.7); (Found: C, 74.86; H, 10.71. C<sub>21</sub>H<sub>36</sub>O<sub>3</sub> requires C, 74.95; H, 10.78%). <sup>1</sup>H NMR:  $\delta$ 0.60 (Me-20), 0.85 (t, Me-15), 1.17 (Me-18), 1.19 (Me-16), 1.29 (Me-17), and 3.63 (OMe). EIMS m/z (rel. int.): 336 (M<sup>+</sup>, 1), 321 (22), 307 (100), 289 (50), 275 (21), 261 (15), 247 (51), 236 (12), 229 (43), 189 (12), 139 (28), 121 (73), 109 (32), and 107 (39) (cf. MS for dihydromanoyl oxide [22]).

The acids in selected mother liquors from the crystallization of 1a were methylated with CH<sub>2</sub>N<sub>2</sub> and chromatographed on 20% AgNO<sub>3</sub>-SiO<sub>2</sub> with Et<sub>2</sub>O-petrol. The first major compound to elute (10% Et<sub>2</sub>O-petrol) was recrystallized twice from pentane to give pure 2b: mp 88-88.5°,  $[\alpha]_D^{25}$  -61° (c0.7); (Found: C, 75.18; H, 10.07. C<sub>21</sub>H<sub>34</sub>O<sub>3</sub> requires C, 75.33; H, 10.23%). <sup>1</sup>H NMR:  $\delta 0.56$  (Me-20), 1.17 (Me-18), 1.23 (Me-16), 1.14 (Me-17), 3.62 (OMe), 4.91 (1H, dd, J = 0.5 and 10.8 Hz), 4.96 (1H, dd, J = 0.5and 18 Hz) and 6.01 (1H, dd, J = 10.8 and 18 Hz). The <sup>13</sup>C NMR is given in Table 1. The MS of 2b is indistinguishable from that of 1b. The IR spectrum (in CCl<sub>4</sub> for 4000-1350 cm<sup>-1</sup> and CS<sub>2</sub> for 1350-430 cm<sup>-1</sup> per the requirements of the Coblentz Society for Class II spectra [23]) of 2b is nearly identical to that of 1b with the exception of vinyl CH2 out-of-plane deformation at 910 cm<sup>-1</sup> for 2b and 920 cm<sup>-1</sup> for 1b (a similar difference between axial and equatorial vinyl CH2 deformation is seen for methyl pimarate and sandaracopimarate [24]) and 'fingerprint' differences between 1250 and 1050 cm<sup>-1</sup>.

Manoyl oxide acid methyl ester 1b eluted from AgNO<sub>3</sub>-SiO<sub>2</sub> after 2b, and was closely followed by methyl imbricataloate 4 (identified by GC retention characteristics and MS, NMR, and IR spectra [13, 24]). Increasing the Et<sub>2</sub>O content of the eluant to 20% gave methyl communate [3], identified by spectral (NMR, UV, IR and MS) and physical data (GC retention and optical rotation) [24, 25].

GC determination of resin acid composition. Fresh needles were cut into ca 1 cm pieces and extracted 5 times with Et<sub>2</sub>O. The acidic fraction of the extract, consisting almost entirely of diterpene resin acids, was quantitatively separated using DEAE-Sephadex [26, 27]. The acids (in 9:1 Et<sub>2</sub>O-MeOH) were converted to the methyl esters with ethereal diazomethane. Composition of the resin acids was determined by GC with DEGS and SE-30/EGiP packed columns [14] at 186° (Table 2).

Retention data with capillary columns has been reported by Foster and Zinkel [28].

Acknowledgements—We are grateful to Dr D. T. Lester, formerly of the University of Wisconsin, for providing provenance and seedling samples of *P. resinosa* needles, and to Dr W. B. Critchfield, Pacific Southwest Forest Experiment Station, Forest Service, U.S. Department of Agriculture, for needles of the *P. nigra* × resinosa and other crosses.

#### REFERENCES

- Guy, C. (1974) The Weymontaching Birchbark Canoe. Anthropological Papers No. 20, National Museums of Canada, Ottawa.
- 2. Sato, A. and Rudloff, E. V. (1964) Can. J. Chem. 42, 635.
- 3. Wroblewska, H., Conner, A. H. and Rowe, J. W. (1977) Wood Sci. 10, 1.

- 4. Wolter, K. and Zinkel, D. F. (1984) Can. J. For. Res. 14, 452.
- 5. Tobolski, J. J. and Zinkel, D. F. (1982) Forest Sci. 28, 785.
- Zinkel, D. F. and Magee, T. V. (1985) Phytochemistry 24, 1273.
- 7. Bardyshev, I. I., Degtyarenko, A. S. and Pekh, T. I. (1982) Khim. Prir. Soedin. 480.
- 8. Jeffries, P. R. and Payne, T. G. (1965) Aust. J. Chem. 18, 1441.
- Carman, R. M., Craig, W. J. and Shaw, I. M. (1973) Aust. J. Chem. 26, 209.
- Joye, N. M., Jr., Roberts, E. M., Lawrence, R. V., Gough, L. J., Soffer, M. D. and Korman, O. (1965) J. Org. Chem. 30, 429.
- Joye, N. M., Jr. and Lawrence, R. V. (1967) J. Chem. Eng. Data 12, 279.
- 12. Santamour, F. S., Jr. and Zinkel, D. F. (1976) Proceedings 3rd Northeastern Forest Tree Improvement Conference, 52.
- 13. Spalding, B. P., Zinkel, D. F. and Roberts, D. R. (1971) Phytochemistry 10, 3289.
- Nestler, F. H. M. and Zinkel, D. F. (1967) Analyt. Chem. 39, 1118.
- Wright, J. W., Read, R. A., Lester, D. T., Merritt, C. and Mohn, C. (1972) Silvae Genetica 21, 205.
- 16. Fowler, D. P. and Lester, D. T. (1970) Genetics of Red Pine.

- USDA Forest Service Research Paper WO-8. U.S. Department of Agriculture, Washington, D.C.
- 17. Fowler, D. P. and Morris, R. W. (1977) Can. J. For. Res. 7, 343
- 18. Critchfield, W. B. (1963) Silvae Genetica 12, 187.
- Gambles, R. L. and Dengler, R. E. (1982) Can. J. Botany 60, 2788.
- 20. Zinkel, D. F. (1976) Phytochemistry 15, 1073.
- Morris, R. W., Critchfield, W. B. and Fowler, D. P. (1980)
   Silvae Genetica 29, 93.
- 22. Enzell, C. R. and Ryhage, R. (1965) Arkiv Kemi 23, 367.
- 23. Coblentz Society (1982) Deskbook of Infrared Spectra (Craver, C. D., ed.) 2nd edn, pp. 11-20.
- Zinkel, D. F., Zank, L. C. and Wesolowski, M. F. (1971)
   Diterpene Resin Acids—A Compilation of Infrared, Mass,
   Nuclear Magnetic Resonance, Ultraviolet Spectra and Gas
   Chromatographic Retention Data. Forest Products
   Laboratory, Madison, Wis.
- 25. Norin, T. (1965) Acta Chem. Scand. 19, 1020.
- 26. Zinkel, D. F. and Rowe, J. W. (1964) Analyt. Chem. 36, 1160.
- 27. Zinkel, D. F. (1983) J. Wood Chem. Tech. 3, 131.
- 28. Foster, D. O. and Zinkel, D. F. (1982) J. Chromatogr. 248, 89.